Veterinary
Services

National Poultry Improvement Plan

Proposed Changes To Be

Considered At The National Plan Conference

Nashville, Tennessee

June 30-July 2, 1996

61 Years of Poultry Improvement

NATIONAL POULTRY IMPROVEMENT PLAN

PROPOSED CHANGES IN THE NATIONAL POULTRY IMPROVEMENT PLAN

INTRODUCTORY STATEMENT

Present provisions of the National Poultry Improvement Plan are contained in the U.S. Department of Agriculture publication, "National Poultry Improvement Plan and Auxiliary Provisions," APHIS 91-55-021, June 1994 and in Title 9 CFR parts 145 and 147.

The detailed procedure for making changes in the Plan is described in the auxiliary provisions, sections 147.41 through 147.48. Copies of the "National Poultry Improvement Plan and Auxiliary Provisions" are available from each Official State Agency or from the National Poultry Improvement Plan staff, Animal and Plant Health Inspection Service, Veterinary Services, Suite A-102, 1500 Klondike Road, Conyers, Georgia 30207.

Proposed changes and supporting statements in this publication were submitted as provided in section 147.44. They are compiled in this publication for consideration at the 1996 National Plan Conference. This publication is distributed well in advance of the conference so that participants and other interested persons may review the proposed changes and inform conference delegates of their wishes regarding the proposals.

Some proposed changes have a line drawn through a portion of the words while other portions are underscored. The line through the words indicate that they are part of the present provision but would be deleted if the proposal were adopted. The underscored words are the proposed additions to that provision.

Each State is entitled to one official delegate for each of the subparts, B, C, D, E, and F of part 145 in which it has one or more participants at the time of the conference with the exception of subpart F. Since subpart F was established by the General Conference Committee on an interim basis and there are no provisions for subpart F at the printing of this document, each participating State is invited to appoint a delegate for subpart F deliberations. Each delegate will act on proposals affecting the provisions of the program which he represents. For reference purposes, delegates are designated as follows:

subpart B delegates - representing egg-type chickens.
subpart C delegates - representing meat-type chickens.
subpart D delegates - representing turkeys.
subpart E delegates - representing waterfowl, exhibition poultry, and game birds.
subpart F delegates - ostrich.

This compilation of proposed changes includes, in the margin adjacent to the section reference for each proposal, the delegate entitled to vote on the proposal. Some of the changes proposed apply equally to all participants in which case conference action will be determined by the combined vote of all delegates.

Section numbers shown with each proposal refer to the numerical identification of affected provisions as published in "National Poultry Improvement Plan and Auxiliary Provisions," APHIS 91-55-021, June 1994. A review of the listing of section numbers and titles may help in locating the provisions involved.

PROPOSED CHANGES
IN THE
NATIONAL POULTRY IMPROVEMENT PLAN

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Proposal No. 1

Delegates:
B,C,D,E

§§145.23 (b) is amended as follows:

(b) U.S. Pullorum-Typhoid Clean. A flock in which freedom from pullorum and typhoid has been demonstrated to the Official State Agency under the criteria in one of the following paragraphs (b)(1) through (5) of this section: Provided, That a flock qualifying by means of a blood test shall be tested within the past 12 months, except that the retesting of a participating flock which is retained for more than 12 months shall be at least 2 weeks prior to molt. ~~the discretion of the Official State Agency with the concurrence of the Service.~~

Reason: Many breeding flocks are molted and recycled for a variety of reasons. It was felt that flocks should be retested prior to the molting process as a means of clarifying the recertification protocol.

Proponent: Dr. Keith Friendshuh
Minnesota Board of Animal Health
St. Paul, Minnesota

Proposal No. 1

Delegates:
B,C,D §§145.23,145.33,145.43 (b) is amended as follows:

(b) **U.S. Pullorum-Typhoid Clean.** A flock in which freedom from pullorum and typhoid has been demonstrated to the Official State Agency under the criteria in one of the following paragraphs (b)(1) through (5) of this section: Provided, That a flock qualifying by means of a blood test shall be tested within the past 12 months, except that the retesting of a participating flock which is retained for more than 12 months shall be tested at least 2 weeks prior to molt. ~~the discretion of the Official State Agency with the concurrence of the Service.~~

Reason: Many breeding flocks are molted and recycled for a variety of reasons. It was felt that flocks should be retested prior to the molting process as a means of clarifying the recertification protocol.

Proponent: Dr. Keith Friendshuh
Minnesota Board of Animal Health
St. Paul, Minnesota

Proposal No. 2

Delegates:

B,C,E §147.11(a)(2) This proposal adds a selenite broth as a selective enrichment broth in the laboratory procedure recommended for the bacteriological cultural examination of environmental monitoring programs and in the pullorum-typhoid reactor bacteriological cultural examination for salmonella.

(2) Selective enrichment culture (refer to illustration 2). Collect and culture organ samples separately from intestinal samples, with intestinal tissues collected last to prevent cross-contamination. Samples from the following organs or sites should be collected for culture in ~~selective enrichment broth~~ both tetrathionate brilliant green (TGB) (Hajna or Mueller-Kauffman) broth and a selenite broth. A non-selective broth culture (illustration 1) of pooled organs and sites should also be included as described in paragraph (a)(3) of this section.

(i) Heart (apex, pericardial sac, and contents if present.);
(ii) Liver (portions exhibiting lesions or in grossly normal organs, the drained gallbladder and adjacent liver tissues.);
(iii) Ovary-Testes (entire inactive ovary or testes, but if ovary is active, include any atypical ova.);
(iv) Oviduct (if active, include any debris and dehydrated ova.);
(v) Kidneys and spleen; and
(vi) Other visible pathological sites where purulent, necrotic, or proliferative lesions are seen.

(3) From each reactor, aseptically collect 10 to 15 g, or the nearest lesser amount available, from each organ or site listed in paragraph (a)(2) of this section and mince, grind, and blend them completely in 10 times their volume of beef extract broth or a comparable non-selective broth. Organs or sites listed in paragraph (a)(2) of this section may be pooled from the same individual bird. Suspensions should be transferred in 10-ml aliquots to 100ml of ~~both~~ tetrathionate brilliant green (TBG) (Hajna or Mueller-Kauffman) broth, a selenite broth, and a separate non-selective broth and incubated 37°C for 24 hours. Refer to illustration 2 for recommended bacteriological recovery and identification procedures, *including delayed secondary enrichment* and combinations of plating media that significantly suppress the overgrowth of contaminants, such as brilliant green Novobiocin (BGN) and Xylose-Lysine-Tergitol 4 (XLT4).

(4) From each reactor, make a composite sample of the following parts of grossly normal or disease tissues from the digestive tract: Crop wall, duodenum (including portions of the pancreas), jejunum (including remnant of yolk-sac attachment), both ceca, cecal tonsils, and rectum-cloaca. Aseptically collect 10-15 g or the nearest lesser amount available from each specified digestive or intestinal tissue, and mince, grind, and blend them completely in 10 times their volume of ~~TBG~~ beef extract broth. The digestive/intestinal tissues may be pooled from the same individual bird. Do not pool tissues from different birds. Transfer 10 ml of the described digestive ~~TBG~~ suspensions into 100 ml of both TBG and a selenite broth, and incubate at 41.5°C for 24 hours. Cultures may be incubated at 37°C if 41.5°C incubators are not available. The higher incubation temperatures for TBG broth reduce populations of competitive contaminants common in gut tissue. Refer to illustration 2 for recommended bacteriological recovery and identification procedures, *including delayed secondary enrichment* and combinations of plating media that significantly suppress the overgrowth of contaminants, such BGN and XLT4.

Illustration 2. Footnote 2 Hajna TT or Mueller-Kauffmann tetrathionate ~~enrichment broth is preferred over selenites~~ and a selenite enrichment broth.

Reason: Experimental studies with an atypical Salmonella pullorum/gallinarum isolate revealed an increased recovery rate for the organism in a selenite enrichment broth when paired organ samples were cultured in both tetrathionate and selenite enrichment broths at varying times post-inoculation.

Proponents: Drs. June degraft-Hanson and George Stein, Jr.
Maryland Department of Agriculture

Proposal No. 3

Delegates:

B §145.23(d) is amended as follows:

(d) U.S. S. Enteritidis Monitored. This program is intended to be the basis from which the breeding-hatching industry may conduct a program for the prevention and control of Salmonellosis. It is intended to reduce the incidence of Salmonella organisms in hatching eggs and chicks through an effective and practical sanitation program at the breeder farm and in the hatchery. This will afford other segments of the poultry industry an opportunity to reduce the incidence of Salmonella in their products.

(1) A flock and the hatching eggs and chicks produced from it which have met the following requirements as determined by the Official State Agency:

(i) The flock originated from a U.S. Sanitation Monitored flock, or meconium from the chick boxes and a sample of chicks that died within 7 days after hatching are examined bacteriologically for salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped.

(ii) All feed fed to the flock shall meet the following requirements:

(A) Pelletized feed shall contain either no animal protein or only animal protein products produced under the Animal Protein Products Industry (APPI) Salmonella Education/Reduction Program[1]. The protein products must have a minimum moisture content of 14.5 percent and must have been heated throughout to a minimum temperature of 190 °F, or above, or to a minimum temperature of 165 °F. for at least 20 minutes, or to a minimum temperature of 184 °F. under 70 lbs. pressure during the manufacturing process.

(B) Mash feed shall contain either no animal protein or only animal protein products supplement manufactured in pellet form and crumbled.

(iii) Feed shall be stored and transported in such a manner as to prevent possible contamination;

(iv) The flock is maintained in compliance with §§ 147.21, 147.24(a), and 147.26 of this chapter;

(v) Environmental samples shall be collected from the flock by an Authorized Agent, as described in § 147.12 of this chapter, when the flock is <u>4 months of age: *Provided*, that multiplier breeding flocks which are to be vaccinated with a federally licensed SE bacterin should be sampled at</u> 2 to 4 weeks of age. The Authorized Agent shall also collect samples every 30 days after the first sample has been collected. The samples shall be examined bacteriologically for group D salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped.

(vi) A federally licensed *Salmonella enteritidis* bacterin may be used in multiplier breeding flocks that are negative for *Salmonella enteritidis* upon bacteriological examination as described in paragraph (d)(1)(v) of this section: *Provided*, that a sample of 350 birds, which will be banded for identification, shall remain unvaccinated until the flock reaches at least 4 months of age. Following negative serological and bacteriological examinations as described in paragraph (d)(1)(vii) of this section, the banded, non-vaccinated birds shall be vaccinated.

(vii) Blood samples from 300 non-vaccinated birds as described in paragraph (d)(1)(vi) of this section shall be officially tested with pullorum- typhoid antigen when the flock is a minimum of more than 4 months of age. All birds with positive or inconclusive reactions, up to a maximum of 25 birds, shall be submitted to an authorized laboratory and examined for the presence of group D salmonella, as described in § 147.11 of this chapter. Cultures from positive samples shall be serotyped.

(viii) Hatching eggs are collected as quickly as possible and are handled as described in § 147.22 of this chapter and are sanitized or fumigated (see § 147.25 of this chapter).

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[1]Documents concerning the APPI/Salmonella Education Reduction Program may be obtained from Mr. A. R. Rhorer; National Animal Health Programs Staff; VS, APHIS, USDA; Suite A-102, 1500 Klondike Rd, Conyers, Georgia, 30207.

(ix) Hatching eggs produced by the flock are incubated in a hatchery that is in compliance with the recommendations in §§ 147.23 and 147.24(b) of this chapter, and sanitized either by a procedure approved by the Official State Agency or fumigated (see § 147.25 of this chapter).
(2) A flock shall not be eligible for this classification if *Salmonella enteritidis* ser enteritidis (SE) is isolated from a specimen taken from a bird in the flock. Isolation of SE from an environmental or other specimen as described in section (d)(1)(v) of this paragraph will require bacteriological examination for SE in an authorized laboratory, as described in § 147.11(a) of this chapter, of a random sample of 60 live birds from a flock of 5,000 birds or more, or 30 live birds from a flock with fewer than 5,000 birds. If only one specimen is found positive for SE, the participant may request bacteriological examination of a second sample, equal in size to the first sample, from the flock. If no SE is recovered from any of the specimens in the second sample, the flock will be eligible for the classification.
(3) A flock shall be eligible for this classification if *Salmonella enteritidis* (*S. enteritidis* ser Enteritidis) is isolated from an environmental sample collected from the flock in accordance with paragraph (d)(v) of this section: *Provided*, That testing is conducted in accordance with paragraph (d)(1)(vi) of this section each 30 days and no positive samples are found.
(4) In order for a hatchery to sell products of this classification, all products handled shall meet the requirements of the classification.
(5) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures.

Reason: Testing the environment of multiplier breeding flocks at 2 to 4 weeks of age requires involving the pullet grow-out facility. Many of the pullet flocks are in different States than the production facility. This early testing was to respond to vaccinated multiplier breeding flocks. The percentage of multiplier flocks that are vaccinating with a federally licensed bacterin for SE is quite small.

Proponent: Mr. Jack Heavenridge
Ohio Poultry Association
Columbus, Ohio

Proposal No. 4

Delegates:

C §145.33(i) Adds a salmonella monitored program for primary meat breeders.

(i) U.S. Salmonella Monitored. This program is intended to be the basis from which the breeding-hatching industry may conduct a program for the prevention and control of Salmonellosis. It is intended to reduce the incidence of Salmonella organisms in hatching eggs and chicks through an effective and practical sanitation program at the breeder farm and in the hatchery. This will afford other segments of the poultry industry an opportunity to reduce the incidence of Salmonella in their products.

(1) A flock and the hatching eggs and chicks produced from it which have met the following requirements as determined by the Official State Agency:

(i) The flock shall originate from a source where sanitation and management practices, as outlined in § 145.33(d)(1) of this paragraph, are conducted;

(ii) The flock is maintained in compliance with §§ 147.21, 147.24(a), and 147.26 of this chapter;

(iii) If pelletized feed contains animal protein, the protein products should be purchased from participants in the Animal Protein Products Industry (APPI) Salmonella Education/Reduction Program. The protein products must have a minimum moisture content of 14.5 percent and must have been heated throughout to a minimum temperature of 190 °F. or above, or to a minimum temperature of 165 °F. for at least 20 minutes, or to a minimum temperature of 184 °F. under 70 lbs. pressure during the manufacturing process;

(iv) If mash feed contains animal protein, the protein products should be purchased from participants in the Animal Protein Products Industry (APPI) Salmonella Education/Reduction Program;

(v) Feed shall be stored and transported in such a manner as to prevent possible contamination;

(vi) Chicks shall be hatched in a hatchery meeting the requirements of §§ 147.23 and 147.24(b) and sanitized or fumigated (see § 147.25 of this chapter) An Authorized Agent shall take environmental samples from the hatchery every 30 days; i.e., (egg shell fragments, meconium, chick papers). An authorized laboratory for Salmonella shall examine the samples bacteriologically;

(vii) An Authorized Agent shall take environmental samples as described in § 147.12 of this chapter, from each flock at 4 months of age and every 30 days thereafter. An authorized laboratory for Salmonella shall examine the environmental samples bacteriologically;

(viii) Owners of flocks found infected with a paratyphoid Salmonella may vaccinate these flocks: Provided, That a sample of 350 birds, which will be banded for identification, shall remain unvaccinated until the flock reaches at least 4 months of age. Following negative serological and bacteriological examinations, the banded, non-vaccinated birds shall be vaccinated.

(2) The Official State Agency may use the procedures described in § 147.14 of this chapter to monitor the effectiveness of the egg sanitation practices.

(3) In order for a hatchery to sell products of this classification, all products handled shall meet the requirements of the classification.

(4) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures.

Reason:
This change provides a vehicle for primary breeders to achieve a classification that will facilitate attaining the export requirements of many foreign customers.

Dr. Louis van der Heide
University of Connecticut
Storrs, Connecticut

Proposal No. 5

Delegates:
C §145.33(j) Establishes a U.S. *Mycoplasma gallisepticum* Monitored program for multiplier breeders.

§ 145.33(j) U.S. Mycoplasma gallisepticum Monitored. It is a flock in which all birds or a sample of at least 20 birds per house has been tested for M. gallisepticum as provided in §145.14(b) when more than 4 months of age: Provided, That to retain this classification, a minimum of 20 birds per house shall be tested at intervals of not more than 90 days: And provided further, That the 20 bird sample should come from two locations within the house (ten from the front half of the house and ten from the back half of the house). A representative sample of males and females should be sampled. The samples should be marked male and female.
(2) A participant handling U.S. M. Gallisepticum Monitored products shall keep these products separate from other products in a manner satisfactory to the Official State Agency: Provided, That U.S. M. Gallisepticum Monitored chicks from primary breeding flocks shall be produced in incubators and hatchers in which only eggs from flocks qualified under paragraph (c)(1)(i) of this section are set.
(3) U.S. M. Gallisepticum Monitored chicks shall be boxed in clean boxes and delivered in trucks that have been cleaned and disinfected as described in § 147.24(a) of this chapter.

Reason:
We feel as though there are potential problems lurking in the multiplier breeder population. Since many multiplier breeders do not participate in the U.S. MG and MS Clean program, there is no structured program for timely bloodtests. Even in the U.S. MG and MS Clean programs there are no guidelines for how to collect the samples or for collecting any samples from males. We feel it is more important to collect a few samples from several locations rather than a large number from one location. Also, most of our positives have originated in our male population after maturity and peak production when the birds are under the most stress. We would still allow a multiplier company to participate in the U.S. MG and MS Clean Program if it chooses to collect the extra samples, but we would ask that all multipliers at least participate in the U.S. MG and MS monitored program. This would allow us to monitor all flocks in the State on a routine basis and be fairer to all companies when we report positive flocks.

Proponent:
Dr. J. Lee Alley
State Veterinarian
Montgomery, Alabama

Proposal No. 6

Delegates:

C §145.33(c) Amends the MG Clean program as follows:

(c) U.S. M. Gallisepticum Clean. (1) A flock maintained in compliance with the provisions of § 147.26 of this chapter and in which freedom from *M. gallisepticum* has been demonstrated under the criteria specified in paragraph (c)(1) (i) or (ii) of this section.

(i) It is a flock in which all birds or a simple of at least 300 birds has been tested for *M. gallisepticum* as provided in § 145.14(b) when more than 4 months of age: Provided, That to retain this classification, a minimum of 150 birds shall be tested at intervals of not more than 90 days: And provided further, That a sample comprised of less than 150 birds may be tested at any one time, with the approval of the Official State Agency and the concurrence of the Service, provided that a minimum of 150 birds is tested within each 90-day period; or

(ii) It is a multiplier breeding flock which originated as U.S. M. Gallisepticum Clean chicks from primary breeding flocks and from which a sample comprised of a minimum of 150 birds per flock has been tested for *M. gallisepticum* as provided in §145.14(b) when more than 4 months of age: Provided, That to retain this classification, the flock shall be subjected to one of the following procedures:

(A) At intervals of not more than 90 days, a sample of 75 birds, with a minimum of 30 birds per pen, whichever is greater shall be tested; or

(B) At intervals of not more than 30 days, a sample of 25 cull chicks produced from the flock shall be subjected to laboratory procedures acceptable to the Official State Agency and approved by the Service, for the detection and recovery of *M. gallisepticum*; or

(C) At intervals of not more than 30 days, egg yolk testing shall be conducted in accordance with § 147.8.

(D) When augmenting male fertility with the addition of male breeding birds, test 3% of males being moved with a minimum of 10 tested per pen. They must be tested between 5 and 10 days of moving. Can be tested by serology, PCR technique, or both methods. The lab can pool 5 swabs per test on the PCR technique. If HI's of 1:40 or PCR positives are found, the males cannot be moved. They must be either destroyed or retested.

(2) A participant handling U.S. M. Gallisepticum Clean products shall keep these products separate from other products in a manner satisfactory to the Official State Agency: Provided, That U.S. M. Gallisepticum Clean chicks from primary breeding flocks shall be produced in incubators and hatchers in which only eggs from flocks qualified under paragraph (c)(1)(i) of this section are set.

(3) U.S. M. Gallisepticum Clean chicks shall be boxed in clean boxes and delivered in trucks that have been cleaned and disinfected as described in § 147.24(a) of this chapter.

Reason:
We also feel that there is a need for a testing program for the use of spike males that many of the multiplier companies use today. We have discussed this with our industry and think that we have developed a program that will be accepted by industry and will aid in the prevention of MG and MS being introduced into our breeder flocks by spike males.

Proponent:
Dr. J. Lee Alley
State Veterinarian
Montgomery, Alabama

Proposal No. 7

Delegates:

C §145.33(k) Establishes a U.S. *Mycoplasma synoviae* Monitored program for multiplier breeders.

It is a flock in which all birds or a sample of at least 20 birds per house has been tested for M. gallisepticum as provided in §145.14(b) when more than 4 months of age: Provided, That to retain this classification, a minimum of 20 birds per house shall be tested at intervals of not more than 90 days: And provided further, That the 20 bird sample should come from two locations within the house (ten from the front half of the house and ten from the back half of the house). A representative sample of males and females should be sampled. The samples should be marked male and female.

(2) A participant handling U.S. M. synoviae Monitored products shall keep these products separate from other products in a manner satisfactory to the Official State Agency: Provided, That U.S. M. synoviae Monitored chicks from primary breeding flocks shall be produced in incubators and hatchers in which only eggs from flocks qualified under paragraph (c)(1)(i) of this section are set.

(3) U.S. M. synoviae Monitored chicks shall be boxed in clean boxes and delivered in trucks that have been cleaned and disinfected as described in § 147.24(a) of this chapter.

Reason:

We feel as though there are potential problems lurking in the multiplier breeder population. Since many multiplier breeders do not participate in the U.S. MG and MS Clean program, there is no structured program for timely bloodtests. Even in the U.S. MG and MS Clean programs there are no guidelines for how to collect the samples or for collecting any samples from males. We feel it is more important to collect a few samples from several locations rather than a large number from one location. Also, most of our positives have originated in our male population after maturity and peak production when the birds are under the most stress. We would still allow a multiplier company to participate in the U.S. MG and MS Clean Program if it chooses to collect the extra samples, but we would ask that all multipliers at least participate in the U.S. MG and MS monitored program. This would allow us to monitor all flocks in the State on a routine basis and be fairer to all companies when we report positive flocks.

Proponent:

Dr. J. Lee Alley
State Veterinarian
Montgomery, Alabama

Proposal No. 8

Delegates:

C §145.33(c) Amends the MS Clean program as follows:

(c) U.S. M. synoviae Clean. (1) A flock maintained in compliance with the provisions of § 147.26 of this chapter and in which freedom from *M. synoviae* has been demonstrated under the criteria specified in paragraph (c)(1) (i) or (ii) of this section.

(i) It is a flock in which all birds or a simple of at least 300 birds has been tested for *M. synoviae* as provided in § 145.14(b) when more than 4 months of age: Provided, That to retain this classification, a minimum of 150 birds shall be tested at intervals of not more than 90 days: And provided further, That a sample comprised of less than 150 birds may be tested at any one time, with the approval of the Official State Agency and the concurrence of the Service, provided that a minimum of 150 birds is tested within each 90-day period; or

(ii) It is a multiplier breeding flock which originated as U.S. *M. synoviae* Clean chicks from primary breeding flocks and from which a sample comprised of a minimum of 150 birds per flock has been tested for *M. synoviae* as provided in §145.14(b) when more than 4 months of age: Provided, That to retain this classification, the flock shall be subjected to one of the following procedures:

(A) At intervals of not more than 90 days, a sample of 75 birds, with a minimum of 30 birds per pen, whichever is greater shall be tested; or

(B) At intervals of not more than 30 days, a sample of 25 cull chicks produced from the flock shall be subjected to laboratory procedures acceptable to the Official State Agency and approved by the Service, for the detection and recovery of *M. synoviae*; or

(C) At intervals of not more than 30 days, egg yolk testing shall be conducted in accordance with § 147.8.

(D) When augmenting male fertility with the addition of male breeding birds, test 3% of males being moved with a minimum of 10 tested per pen. They must be tested between 5 and 10 days of moving. Can be tested by serology, PCR technique, or both methods. The lab can pool 5 swabs per test on the PCR technique. If HI's of 1:40 or PCR positives are found, the males cannot be moved. They must be either destroyed or retested.

(2) A participant handling U.S. *M. synoviae* Clean products shall keep these products separate from other products in a manner satisfactory to the Official State Agency: Provided, That U.S. *M. synoviae* Clean chicks from primary breeding flocks shall be produced in incubators and hatchers in which only eggs from flocks qualified under paragraph (c)(1)(i) of this section are set.

(3) U.S. *M. synoviae* Clean chicks shall be boxed in clean boxes and delivered in trucks that have been cleaned and disinfected as described in § 147.24(a) of this chapter.

Reason:

We feel that there is a need for a testing program for the use of spike males that many of the multiplier companies use today. We have discussed this with our industry and think that we have developed a program that will be accepted by industry and will aid in the prevention of MG and MS being introduced into our breeder flocks by spike males.

Proponent:

Dr. J. Lee Alley
State Veterinarian
Montgomery, Alabama

Proposal No. 9

Delegates

B.C,D,E,F — §145.5(c) Provides for exemption of the Ostrich from the P-T Clean State classification.

§145.5(c) A flock shall be deemed to be a participating flock at any time only if it has qualified for the U.S. Pullorum-Typhoid Clean classification, as prescribed in Subparts B, C, D, or E of this part: Provided, that Subpart F (Ostrich) requirements for Pullorum-Typhoid clean is at the discretion of the Official State Agency.

Reason: The General Conference Committee has approved on an interim basis the addition of the Ostrich as Subpart F of the Provisions of the National Poultry Improvement Plan. Many States require that the Ostrich be tested pullorum-typhoid free prior to entering the respective State, however many do not. This change would allow those States who currently have P-T Clean State status to retain it without requiring all ostrich breeding flocks be tested free. The issue is complicated by confounding factors; i.e., 1) the requirement that reactors to the serological test for pullorum-typhoid must be sacrificed for bacteriological culturing to determine the status of the bird; 2) the reagents used to conduct the serological tests for pullorum-typhoid are based on data collected relative to chickens and turkeys therefore may not be as reliable for the ostrich; 3) obviously the age of sexual maturity for the ostrich is different from the chicken or turkey and may well be different between breeds within the same specie.

Proponent:
General Conference Committee

Proposal No. 10

Delegates
B,C,D,E,F §145.10 Terminology and Classification;Flocks, Products, and States.

(c) A flock shall be deemed to be a participating flock, products produced from them, and States which have met the respective requirements specified in Part 145 Subpart B,C,D,E,or F may be designated by the following terms or illustrative designs:
(d) Each bird shall be identified with a sealed and numbered band obtained through or approved by the Official State Agency: Provided, That exception may be made at the discretion of the Official State Agency.

Reason:
The General Conference Committee has approved the addition of the Subpart F (Ostrich) to the provisions of the NPIP. This proposed change is required to enable the ostrich breeder or the State to use Official terminology and/or illustrative designs of the NPIP on any Official NPIP document.

Proponent:
General Conference Committee

Proposal No. 11

Delegates:
B,C,D,E,F **§ 145.14 Blood testing.**

Poultry must be more than 4 months of age when blood tested for an official classification; Provided, That turkey candidates may be blood tested at more than 12 weeks of age under Subpart D, while game birds may be blood tested under Subpart E when more than 4 months of age or upon reaching sexual maturity, whichever comes first; Provided further, that ostrich candidates may be blood tested upon reaching sexual maturity or at the discretion of the Official State Agency. Blood samples for official tests shall be drawn by an Authorized Agent or State Inspector and tested by an authorized laboratory, except that the stained antigen, rapid whole-blood test for pullorum-typhoid may be conducted by an Authorized Agent or State Inspector. For Plan programs in which a representative sample may be tested in lieu of an entire flock, the minimum number tested shall be 30 birds per house, with at least 1 bird taken from each pen and unit in the house. All birds must be tested in houses containing fewer than 30 birds.
(a) For Pullorum-Typhoid. (1) The official blood tests for pullorum-typhoid shall be the standard tube agglutination test, the microagglutination test, the enzyme-labeled immunosorbent assay test (ELISA), or the rapid serum test for all poultry; and the stained antigen, rapid whole-blood test for all poultry except turkeys and ostriches; The procedures for conducting official blood tests are set forth in §§ 147.1, 147.2, 147.3, and 147.5 of this chapter and referenced in footnote 3 of this section or in literature provided by the producer. Only antigens approved by the Department and of the polyvalent type shall be used for the rapid whole-blood and tube agglutination tests. Each serial of tube antigen shall be submitted by the antigen producer to the Department for approval upon manufacture and once a year thereafter as long as antigen from that serial continues to be made available for use. All microtest antigens and enzyme-labeled immunosorbent assay reagents shall also be approved by the Department./1/

(2) [Reserved]
(3) There shall be an interval of at least 21 days between any official blood test and any previous test with pullorum-typhoid antigen.
(4) [Reserved]
(5) The official blood test shall include the testing of a sample of blood from each bird in the flock: Provided, That under specified conditions (see applicable provisions of §§ 145.23, 145.33, 145.43 and 145.53) the testing of a portion or sample of the birds may be used in lieu of testing each bird.
(6) When reactors are found from any flock, or S. pullorum or S. gallinarum organisms are isolated by an authorized laboratory from baby poultry, or from fluff samples produced by hatching eggs, the infected flock shall qualify for participation in the Plan with two consecutive negative results to an official blood test named in paragraph
(a)(1) of this section. A succeeding flock must be qualified for participation in the Plan's pullorum-typhoid program with a negative result to an official blood test named in paragraph (a)(1) of this section. Testing to qualify flocks for Plan participation must include the testing of all birds in infected and succeeding flocks for a twelve month period, and shall be performed or physically supervised by a State Inspector; Provided, That at the discretion of the Official State Agency, a sample of at least 500 birds, rather than all birds in the flock, may be tested by the State Inspector if it is agreed upon by the Official State Agency, the flockowner, and the Administrator. If the State Inspector determines that a primary breeding flock has been exposed to *S. pullorum* or *S. gallinarum*,/2/ the Official State Agency may require:

(i) The taking of blood samples--performed by or in the presence of a State Inspector--from all birds on premises exposed to birds, equipment, supplies,

or personnel from the primary breeding flock during the period when the State Inspector determined that exposure to *S. pullorum* or *S. gallinarum* occurred./2/
(ii) The banding of all birds on these premises--performed or physically supervised by a State Inspector--in order to identify any bird that tests positive; and
(iii) The testing of blood samples at an authorized laboratory using an official blood test named in paragraph (a)(1) of this section.
(7) All domesticated fowl, except waterfowl, on the farm of the participant shall either be properly tested to meet the same standards as the participating flock or these birds and their eggs shall be separated from the participating flock and its eggs.
(8) All tests for pullorum-typhoid in flocks participating in or candidates for participation in the Plan shall be reported to the Official State Agency within 10 days following the completion of such tests. All reactors shall be considered in determining the classification of the flock.
(9) Poultry from flocks undergoing qualification testing for participation in the Plan, that have a positive reaction to an official blood test named in paragraph (a)(1) of this section, shall be evaluated for pullorum-typhoid infection. The Official State Agency shall select one or more of the following procedures to be used in each circumstance, based on a cost-benefit analysis involving evaluation of such factors as: the value of the reactors and flocks at risk; the necessity for preserving birds from scarce genetic lines; the need for a quick determination of disease existence; and the cost for each retesting option versus the total availability of funds (when the State provides retesting subsidies):
(i) Reactors shall be submitted to an authorized laboratory for bacteriological examination. If there are more than 4 reactors in a flock, a minimum of 4 reactors shall be submitted to the authorized laboratory; if the flock has 4 or fewer reactors, all of the reactors must be submitted. The approved procedure for bacteriological examination is set forth in § 147.11 of this chapter. When reactors are submitted to the authorized laboratory within 10 days from the date of reading an official blood test named in paragraph (a)(1) of this section, and the bacteriological examination fails to demonstrate pullorum-typhoid infection, the Official State Agency shall presume that the flock has no pullorum-typhoid reactors.
(ii) The serum specimen that produced the positive reaction shall be retested at an authorized laboratory in accordance with procedures set forth in § 147.1 of this chapter for the standard tube agglutination test, or in § 147.5 of this chapter for the microagglutination test for pullorum- typhoid. If the reaction to this retest is positive in dilutions of 1:50 or greater for the standard tube agglutination test, or 1:40 or greater for the microagglutination test, additional examination of the bird and flock will be performed in accordance with paragraph (a)(9)(i) or (a)(9)(iii) of this section.
(iii) The reactors shall be retested within 30 days using an official blood test named in paragraph (a)(1) of this section. If this retest is positive, additional examination of the reactors and flock will be performed in accordance with paragraph (a)(9)(i) of this section; Provided, that the Official State Agency has the option of using an approved bacteriological monitoring test in lieu of the blood test for the ostrich. During this 30-day period, the flock must be maintained under a security system, specified or approved by the Official State Agency, that will prevent physical contact with other birds and assure that personnel, equipment, and supplies that could be a source of pullorum-typhoid spread are sanitized.
(10) Any drug, for which there is scientific evidence of masking the test reaction or hindering the bacteriological recovery of Salmonella organisms, shall not be fed or administered to poultry within 3 weeks prior to a test or bacteriological examination upon which a Salmonella classification is based.
(11) When suitable evidence, as determined by the Official State Agency or the State Animal Disease Control Official, indicates that baby or started poultry produced by participating hatcheries are infected with organisms for

which the parent flock received an official control classification and this evidence indicates that the infection was transmitted from the parent flock, the Official State Agency may, at its discretion, require additional testing of the flock involved. If infection is found in the parent flock, its classification shall be suspended until the flock is requalified under the requirements for the classification. Furthermore, the Official State Agency may require that the hatching eggs from such flocks be removed from the incubator and destroyed prior to hatching. When Salmonella organisms are isolated from a specimen which originated in a participating hatchery, the Official State Agency shall attempt to locate the source of the infection. The results of the investigation and the action taken to eliminate the infection shall be reported by the Official State Agency to the Service.

Reason: The General Conference Committee has approved on an interim basis the addition of the Ostrich as Subpart F of the Provisions of the National Poultry Improvement Plan. Many States require that the Ostrich be tested pullorum-typhoid free prior to entering the respective State. The issue is complicated by confounding factors; i.e., 1) the requirement that reactors to the serological test for pullorum-typhoid must be sacrificed for bacteriological culturing to determine the status of the bird; 2) the reagents used to conduct the serological tests for pullorum-typhoid are based on data collected relative to chickens and turkeys therefore may not be as reliable for the ostrich; 3) obviously the age of sexual maturity for the ostrich is different from the chicken or turkey and may well be different between breeds within the same specie.

Proponent:
General Conference Committee

Proposal No. 12

Delegates

F Subpart F--Special Provisions for Ostrich Breeding Flocks and Products

§145.61 Definitions.
Except where the context otherwise requires, for the purposes of this subpart the following terms shall be construed, respectively, to mean:

Chicks Newly hatched ostriches which have not been fed or watered.

§145.42 Participation.

(a) Participating ostrich flocks, and the eggs and chicks produced from them, shall comply with the applicable general provisions of Subpart A of this part and the special provisions of this Subpart F.

(b) The minimum weight of ostrich hatching eggs shipped interstate shall be

(c) Hatching eggs shall be fumigated(see §147.25 of this chapter) or otherwise sanitized.

§ 145.63 Terminology and Classification; Flocks and Products.

Participating flocks, and the eggs and chicks produced from them, which have met the respective requirements specified in this section may be designated by the following terms and the corresponding designs illustrated in § 145.10:
(a) [Reserved]
(b) U.S. Pullorum-Typhoid Clean. A flock in which freedom from pullorum and typhoid has been demonstrated to the Official State Agency under the criteria in one of the following paragraphs (b) (1) through (5) of this section: Provided, That a flock qualifying by means of a blood test shall be tested within the past 12 months, except that the retesting of a participating flock which is retained for more than 12 months shall be at the discretion of the Official State Agency with the concurrence of the Service. (See § 145.14 relating to the official blood test where applicable.)
(1) It has been officially blood tested with no reactors.
(i) The flock is located in a State where all persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarum is isolated;
(ii) The flock is composed entirely of birds that originated from U.S. Pullorum-Typhoid Clean breeding flocks or from flocks that met equivalent requirements under official supervision; and
(iii) The flock is located on a premises where either no poultry or a flock not classified as U.S. Pullorum-Typhoid Clean were located the previous year; Provided, That an Authorized Agent must blood test all birds, as described in § 145.14, if the Official State Agency determines that the flock has been exposed to pullorum-typhoid. In making determinations of exposure and setting the number of birds to be blood tested, the Official State Agency shall evaluate the results of any blood tests, described in § 145.14(a)(1) of this part, that were performed on an unclassified flock located on the premises during the previous year; the origins of the unclassified flock; and the probability of contacts between the flock for which qualification is being sought and (a) infected wild birds, (b) contaminated feed or waste, or (c) birds, equipment, supplies, or personnel from flocks infected with pullorum-typhoid.

(i) All ostrich hatcheries within the State are qualified as "National Plan Hatcheries" or have met equivalent requirements for pullorum-typhoid control

under official supervision;
(ii) All ostrich hatchery supply flocks within the State are qualified as U.S. Pullorum-Typhoid Clean or have met equivalent requirements for pullorum-typhoid control under official supervision: Provided, That if other domesticated fowl, except waterfowl, are maintained on the same premises as the participating flock, freedom from pullorum-typhoid infection shall be demonstrated by an official blood test of each of these fowl;
(iii) All shipments of products other than U.S. Pullorum-Typhoid Clean, or equivalent, into the State are prohibited;
(iv) All persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which S. pullorum or S. gallinarum is isolated;
(v) All reports of any disease outbreak involving a disease covered under the Plan are promptly followed by an investigation by the Official State Agency to determine the origin of the infection; Provided, That if the origin of the infection involves another State, or if there is exposure to poultry in another State from the infected flock, then the National Poultry Improvement Plan will conduct an investigation;
(vi) All flocks found to be infected with pullorum or typhoid are quarantined until marketed or destroyed under the supervision of the Official State Agency, or until subsequently blood tested, following the procedure for reacting flocks as contained in § 145.14(a)(5), and all birds fail to demonstrate pullorum or typhoid infection;
(vii) [Reserved]
(viii) Discontinuation of any of the conditions or procedures described in paragraphs (b)(3) (i), (ii), (iii), (iv), (v), and (vi) of this section, or the occurrence of repeated outbreaks of pullorum or typhoid in ostrich breeding flocks within or originating within the State shall be grounds for the Service to revoke its determination that such conditions and procedures have been met or complied with. Such action shall not be taken until a thorough investigation has been made by the Service and the Official State Agency has been given an opportunity to present its views.
(5) It is a primary breeding flock located in a State determined to be in compliance with the provisions of paragraph (b)(4), of this section and in which each bird in has been officially tested for pullorum-typhoid with no reactors: Provided, That a bacteriological examination monitoring program acceptable to the Official State Agency and approved by the Service may be used in lieu of blood testing.

Reason:
The ostrich industry requested that the General Conference Committee allow on an interim basis the addition of the ostrich as subpart F to the provisions of the NPIP in an attempt to establish more uniform health requirements from State to State for the ostrich. The General Conference Committee approved the establishment of a subpart F for the ostrich.

Proponent:
General Conference Committee

Proposal No. 13

Delegates:
B,C,D,E,F **§ 147.45 Amends the section as follows:**

§ 147.45 Official Delegates.

Each cooperating State shall be entitled to one official delegate for each of the programs prescribed in Subparts B, C, D, ~~and~~, E and F of Part 145 of this chapter in which it has one or more participants at the time of the Conference. The official delegates shall be elected by a representative group of participating industry members and be certified by the Official State Agency. It is recommended but not required that the official delegates be Plan participants. Each official delegate shall endeavor to obtain, prior to the Conference, the recommendations of industry members of his State with respect to each proposed change.

Reason:
The ostrich industry requested that the General Conference Committee allow on an interim basis the addition of the ostrich as subpart F to the provisions of the NPIP in an attempt to establish more uniform health requirements from State to State for the ostrich. The General Conference Committee approved the establishment of a subpart F for the ostrich.

Proponent:
General Conference Committee

Proposal No. 14

Delegates:
B,C,D,E,F **§ 147.46 Committee Consideration of Proposed Changes.**

(a) The following ~~four~~ five committees shall be established to give preliminary consideration to the proposed changes falling in their respective fields:
(1) Egg-type chickens.
(2) Meat-type chickens.
(3) Turkeys.
(4) Waterfowl, exhibition poultry, and game birds.
(5) Ostrich.
(b) Each official delegate shall be appointed a voting member in one of the committees specified in paragraph (a) of this section.
(c) Since several of the proposals may be interrelated, the committees shall consider them as they may relate to others, and feel free to discuss related proposals with other committees.
(d) The committees shall make recommendations to the conference as a whole concerning each proposal. The committee report shall show any proposed change in wording and the record of the vote on each proposal, and suggest an effective date for each proposal recommended for adoption. The individual committee reports shall be submitted to the chairman of the conference, who will combine them into one report showing, in numerical sequence, the committee recommendations on each proposal.
(e) The committee meetings shall be open to any interested person. Advocates for or against any proposal should feel free to appear before the appropriate committee and present their views.

Reason:
The ostrich industry requested that the General Conference Committee allow on an interim basis the addition of the ostrich as subpart F to the provisions of the NPIP in an attempt to establish more uniform health requirements from State to State for the ostrich. The General Conference Committee approved the establishment of a subpart F for the ostrich.

Proponent:
General Conference Committee

Proposal No. 15

Delegates:
F **§ 147.41 Definitions.**

Except where the context otherwise requires, for the purposes of this subpart the following terms shall be construed, respectively, to mean:
Department. The U.S. Department of Agriculture.

Egg-type chickens. Chickens bred for the primary purpose of producing eggs for human consumption.

Exhibition poultry. Domesticated fowl which are bred for the combined purposes of meat or egg production and competitive showing.

Game birds. Domesticated fowl, such as pheasants, partridge, quail, grouse, and guineas, but not doves and pigeons.

Meat-type chickens. Chickens bred for the primary purpose of producing meat.

Ostrich. A swift-footed flightless bird of the genus Struthio having a downy neck and head, thighs nearly bare, and two-toed feet.

Plan Conference. A meeting convened for the purpose of recommending changes in the provisions of the Plan.

Plan or NPIP. The National Poultry Improvement Plan.

Service. The Animal and Plant Health Inspection Service, Veterinary Services, of the Department.

State. Any State, the District of Columbia, or Puerto Rico.

Waterfowl. Domesticated fowl that normally swim, such as ducks and geese.

Reason:
The ostrich industry requested that the General Conference Committee allow on an interim basis the addition of the ostrich as subpart F to the provisions of the NPIP in an attempt to establish more uniform health requirements from State to State for the ostrich. The General Conference Committee approved the establishment of a subpart F for the ostrich.

Proponent:
General Conference Committee

Proposal No. 16

Delegates:
B,C,D,E,F Subpart A—General Provisions

§145.1 Definitions.

Words used in this part in the singular form shall be deemed to import the plural, and vice versa, as the case may demand. Except where the context otherwise requires, for the purposes of this part the following terms shall be construed, respectively, to mean:

Administrator. The Administrator, Animal and Plant Health Inspection Service, or any person authorized to act for the Administrator.

Affiliated flockowner. A flockowner who is participating in the Plan through an agreement with a participating hatchery.

Animal and Plant Health Inspection Service. The Animal and Plant Health Inspection Service of the U.S. Department of Agriculture.

Authorized Agent. Any person designated under § 145.11(a) to perform functions under this part.

Authorized laboratory. A laboratory designated by an Official State Agency, subject to review by the Service, to perform the blood testing and bacteriological examinations provided for in this part.

Baby poultry. Newly hatched poultry (chicks, poults, ducklings, goslings, keets, etc.) that have not been fed or watered.

Colon bacilli. For the purpose of this chapter, those organisms which are gram negative, non spore-forming bacilli, which ferment lactose with gas formation, and serve as an index of fecal contamination.

Dealer. An individual or business that deals in commerce in hatching eggs, newly-hatched poultry, and started poultry obtained from breeding flocks and hatcheries. This does not include an individual or business that deals in commerce in buying and selling poultry for slaughter only.

Department. The U.S. Department of Agriculture.

Domesticated. Propagated and maintained under the control of a person.

Equivalent or equivalent requirements. Requirements which are equal to the program, conditions, criteria, or classifications with which compared, as determined by the Official State Agency and with the concurrence of the Service.

Exposed (Exposure). Contact with birds, equipment, personnel, supplies, or any article infected with, or contaminated by, communicable poultry disease organisms.

Flock--(1) As applied to breeding. All poultry of one kind of mating (breed and variety or combination of stocks) and of one classification on one farm; (2) As applied to disease control. All of the poultry on one farm except that, at the discretion of the Official State Agency, any group of poultry which is segregated from another group and has been so segregated for a period of at least 21 days may be considered as a separate flock.

Fluff sample. Feathers, shell membrane, and other debris resulting from the hatching of poultry.

Fowl typhoid or typhoid. A disease of poultry caused by *Salmonella gallinarum.*
Franchise breeder. A breeder who normally sells products under a specific strain or trade name and who authorizes other hatcheries to produce and sell products under this same strain or trade name.

Franchise hatchery. A hatchery which has been authorized by a franchise breeder to produce and sell products under the breeder's strain or trade name.

Hatchery. Hatchery equipment on one premises operated or controlled by any person for the production of baby poultry.

Infected flock. A flock in which an authorized laboratory has discovered one or more birds infected with a communicable poultry disease for which a program has been established under the Plan.

Midlay. Approximately 2-3 months after a flock begins to lay or after a molted flock is put back into production.

Multiplier breeding flock. A flock that is intended for the production of hatching eggs used for the purpose of producing progeny for commercial egg or meat production or for other non-breeding purposes.

Official State Agency. The State authority recognized by the Department to cooperate in the administration of the Plan.

Official supervision--(1) As applied to Plan programs. The direction, inspection, and critical evaluation by the Official State Agency of compliance with the provisions of the Plan;
(2) As applied to non-Plan but equivalent State poultry improvement programs. The direction, inspection, and critical evaluation by an officer or agency of a State government, of compliance with a publicly announced State poultry improvement program.

Person. A natural person, firm, or corporation.

Plan. The provisions of the National Poultry Improvement Plan contained in this part.

Poultry. Domesticated fowl, including chickens, ostriches, turkeys, waterfowl, and game birds, except doves and pigeons, which are bred for the primary purpose of producing eggs or meat.

Primary breeding flock. A flock composed of one or more generations that is maintained for the purpose of establishing, continuing, or improving parent lines.

Products. Poultry breeding stock and hatching eggs, baby poultry, and started poultry.

Program. Management, sanitation, testing, and monitoring procedures which, if complied with, will qualify, and maintain qualification for, designation of a flock, products produced from the flock, or a state by an official Plan classification and illustrative design, as described in § 145.10 of this part.

Pullorum disease or pullorum. A disease of poultry caused by *Salmonella pullorum.*

Reactor. A bird that has a positive reaction to a test, required or recommended in Parts 145 or 147 of this chapter, for any poultry disease for which a program has been established under the Plan.

Salmonella. Any bacteria belonging to the genus Salmonella, including the

arizona group.

Sanitize. To treat with a product which is registered by the Environmental Protection Agency as germicidal, fungicidal, pseudomonocidal, or tuberculocidal, in accordance with the specifications for use as shown on the label of each product. The Official State Agency, with the concurrence of the Service, shall approve each product or procedure according to its specified usage.

Serial. The total quantity of completed product which has been thoroughly mixed in a single container and identified by a serial number.

Service. The Animal and Plant Health Inspection Service, Veterinary Services, of the Department.

Sexual maturity. The average age at which a species of poultry is biologically capable of reproduction.

Started poultry. Young poultry (chicks, pullets, cockerels, capons, poults, ducklings, goslings, keets, etc.) that have been fed and watered and are less than 6 months of age <u>except for the ostrich</u>.

State. Any State, the District of Columbia, or Puerto Rico.

State Inspector. Any person employed or authorized under § 145.11(b) to perform functions under this part.

Stock. A term used to identify the progeny of a specific breeding combination within a species of poultry. These breeding combinations may include pure strains, strain crosses, breed crosses, or combinations thereof.

Strain. Poultry breeding stock bearing a given name produced by a breeder through at least five generations of closed flock breeding <u>with the exception of the ostrich</u>.

S. typhimurium infection or *typhimurium.* A disease of poultry caused by *Salmonella typhimurium* or *S. typhimurium* var. copenhagen.

Succeeding flock. A flock brought onto a premises during the 12 months following removal of an infected flock.

Suspect Flock. A flock shall be considered, for the purposes of the Plan, to be a suspect flock if any evidence exists that it has been exposed to a communicable poultry disease.

Trade name or number. A name or number compatible with State and Federal laws and regulations applied to a specified stock or product thereof.

Reason:
The ostrich industry requested that the General Conference Committee allow on an interim basis the addition of the ostrich as subpart F to the provisions of the NPIP in an effort to establish more uniform health requirements from State to State for the ostrich. The General Conference Committee approved the establishment of a subpart F for the ostrich.

Proponent:
General Conference Committee

Proposal No. 17

Delegates:
B

§ 145.23 (h) U. S. S. ENTERITIDIS MONITORED STARTED POULTRY. This change would create a program for the prevention and control of Salmonella enteritidis for the started-poultry industry.
(h) **U. S. S. ENTERITIDIS MONITORED STARTED POULTRY**-This program is intended to be the basis from which the started-poultry industry may conduct a program for the prevention and control of Salmonella enteritidis. It is intended to provide reasonable assurance to egg producers that a pullet flock may be certified for a quality assurance S. enteritidis laying flock program.

(h)(1) A pullet flock which has met the following requirements as determined by the Official State Agency:
(h)(1)(i) Replacement chicks shall originate from parent flocks and hatchery which meet the requirements of U.S. Sanitation Monitored for egg type breeders or an equivalent program.
(h)(1)(ii) Baby chicks shall be placed in cleaned and disinfected houses that have been environmentally examined and are negative for Salmonella.
(h)(1)(iii) The flock is maintained in compliance with §§ 147.21, 147.24 (a), and 147.26 of this chapter;
(h)(1)(iv) Environmental samples shall be collected from the grow-out facility by an Authorized Agent, as described in § 147.12 of this chapter, at 7-10 days of age and at 12-18 weeks of age. The samples shall be examined bacteriologically for group D salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped.
(h)(1)(v) Approved rodent, wild bird and fly control programs will be required in the pullet growing facility.
(h)(1)(vi) All feed fed to the flock shall meet one of the following requirements:
(h)(1)(vi)(A) Pelletized feed shall contain either no animal protein or only animal protein products produced under the Animal Protein Products Industry (APPI) Salmonella Education/ Reduction Program, a minimum moisture content of 14.5 percent, and must have been subjected to temperature of 190°F. or above, 165°F. for at least 20 minutes, or 184°F. and 70 lbs. of pressure during the manufacturing process.
(h)(1)(vi)(B) Mash feed shall contain either no animal protein or only animal protein product supplements manufactured in pellet form and crumbled.
(h)(1)(vii) Feed shall be stored and transported in such a manner as to prevent possible contamination;
(h)(1)(viii) A sample of early mortality shall be collected at 7-10 days. The samples shall be examined bacteriologically for group D salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped
(h)(1)(ix) Pullets shall be transported only in cleaned and disinfected crates and vehicles.
(h)(2) Isolation of SE from an environmental or other specimen (early chick mortality) as described in section (h)(1)(iv) or (h)(1)(viii) of this paragraph will require the bacteriological examination of 60 birds from the flock for Salmonella enteritidis in an authorized laboratory as described in section 147.11 of this chapter.
A flock shall not be eligible for this classification if Salmonella enteritidis (S. enteritidis ser Enteritidis) is isolated from the internal organs (heart, liver, gall bladder, spleen, oviduct, and ovary) of a bird in the flock.
(h)(3) A flock shall remain eligible for this classification if Salmonella enteritidis (S. enteritidis ser Enteritidis) is only isolated from the environment.
(h)(4) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures.

REASON:
This is a voluntary program for egg producers or others who wish to have reasonable assurance as to the SE status of pullets which are being added to their laying flocks. This program, although not a program involving breeding flocks or hatching, is very similar to current pullet programs of NPIP which classify pullets in regard to MG and MS.

This program provides documented and approved guidelines for the industry to follow in establishing SE control programs of their own even though not necessarily supervised and classified by an Official State Agency.

Having such a voluntary program available will tend to de-emphasize any perceived need for a mandatory program.

PROPONENT:
Dr. Kenton Kraeger
Hy-Line International
Dallas Center, Iowa

Proposal No. 18

Delegates B,C,D,E,F

§147.25 Amends the section by adding OSHA exposure limits for formaldehyde.

§ 147.25 Fumigation.

Fumigation may be used for sanitizing eggs and hatchery equipment as an essential part of a sanitation program. APHIS disclaims any liability in the use of formaldehyde for failure on the part of the user to adhere to the Occupational Safety and Health Administration (OSHA) standards for formaldehyde fumigation, published in the Dec. 4, 1987, Federal Register (52 FR 46168, Docket Nos. H-225, 225A, and 225B). In addition to extensive safety requirements, OSHA standards relating to the permissible exposure limits are as follows: (1) 1 part per million (PPM) as an 8-hour time weighted average; (2) 2 ppm as a 15 minutes short term exposure limit; and (3) 0.5 ppm action level measured as an 8-hour time weighted average.

Reason:
The hatchery industry needs to know OSHA's standards for permissible exposure limits to employees relative to the use of formaldehyde. The staff of the NPIP felt that providing that information in the provisions of the NPIP would be helpful to the hatchery industry.

Proponent:
Poultry Improvement Staff, National Poultry Improvement Plan.

Proposal No. 19

Delegates:
B,C, §147.12 is amended by adding the procedure for collecting chick papers for bacteriological examination.

§ 147.12 Procedures for Collecting Environmental Samples, chick papers and Cloacal Swabs for Bacteriological Examination.

Information concerning the pen arrangement and number of birds per pen should be obtained from the owner so that the required number of samples per pen and per flock can be determined. A means of identifying each sample by pen of origin should be provided. The vehicle transporting the personnel taking the samples should be left as far as practical from the poultry pens. Sanitary precautions, including personal cleanliness, should be observed during the sampling procedure. The hands should be carefully washed with a sanitizing soap prior to the sampling. Outer clothing, including gloves, should be changed between visits to different premises so that clean clothing is worn upon entering each premises.
The used and clean apparel should be kept separate. Boots or footwear should be cleaned and disinfected between visits to different premises. Disposable caps should be provided and discarded after use on each premises. After collection, the samples should be protected from drying, light, and excessive temperatures and delivered to the laboratory within one day. If delivery is delayed, samples should be refrigerated.
(a) Environmental Samples. Fecal material, litter, or dust to be submitted for bacteriological examination should be collected in accordance with the procedures described in paragraphs (a)(1) or (2) of this section:
(1) Procedure for sampling in broth. Authorized laboratories will provide capped tubes 1-2 cm in diameter and 15-20 cm in length which are two-thirds full of a recently made, refrigerated, sterile enrichment broth (Selenite Brilliant Green Sulfapyridine or Tetrathionate Brilliant Green) for each sample. Sufficient tubes should be taken to the premises to provide at least one tube per pen or one tube per 500 birds, whichever is greater. At least one sterile, cotton-tipped applicator will be needed for each tube. The dry applicator is first placed or drawn through fresh manure (under roost, near water troughs, cecal droppings, or diarrhetic droppings). After this and each subsequent streaking, the cotton-tipped applicator is placed in the tube of broth and swirled to remove the collected material. The applicator is then withdrawn and is used for taking additional specimens by streaking on or through areas where defecation, trampling of feces, or settling of dust are common; i.e., on or near waterers, feeders, nests, or rafters, etc. When the volume of material collected equals approximately 10 percent of the volume of the broth (usually 10-12 streakings), the applicator is placed in the tube and the stick is broken in half. The lower or cotton-tipped half is left in the broth, and the upper half is retained for future disposal. The cap is then replaced on the inoculated tube, and the sampling procedure is continued in other areas of the pen.
(2) Procedure for sampling in dry containers. A sample of fecal material, litter, or dust is placed in a sterile, sealable container. The sample shall consist of several specimens of material taken from a representative location in the pen or house. At least 10 g (approximately a heaping tablespoonful) of material shall be collected for each sample. The specimens in each sample shall be collected with a sterile tongue depressor or similar uncontaminated instrument. The samples should vary in type and consistency. Half of the samples should be comprised of material representing defecated matter from a large portion of the flock; i.e., trampled, caked material near waterers and feeders. The minimum number of samples to be taken shall be determined by the following:

Five samples from pens or houses of up to 500 birds;
Ten samples from pens or houses of 500 to 2,500 birds;

Fifteen samples from pens or houses with more than 2,500 birds.

The composite samples above may be pooled to not less than five samples at the laboratory as long as the volume of material collected equals approximately 10 percent of the volume of the broth.

(b) Cloacal swabs. Cloacal swabs for bacteriological examination are taken from each bird in the flock or from a minimum of 500 birds in accordance with the procedure described in paragraph (a)(1) of this section.

(1) Procedure for taking cloacal swabs. The authorized laboratory will provide sterile capped tubes or other suitable containers and cotton-tipped applicators for use in taking the cloacal swabs. The cotton-tipped applicator is inserted into the cloaca and rectum in such a manner as to insure the collection of fecal material. The swab and adhering fecal material are then placed in the tube and the stick is broken in half, with the upper half retained for future disposal. The cloacal swabs may be combined in the sterile tubes in multiples of five or in combinations specified by the authorized laboratory.

(c) Drag-swabs. Drag-swabs for bacteriological examination should involve the exposure of at least six unpooled pads per house to promote representative sampling and some element of quantification.

(1) Drag-swab assembly. Assemble drag-swab sampling sets from folded-once 3-by-3-inch sterile gauze pads secured with paper clips. Bend end wires of each paper clip slightly to catch into the swab fabric, thus securing the clips to the folded pads. Use two pads, assembled as described to make each drag-swab sampling set. Securely connect one pad through the free rounded end of the paper clip to a 2-ft (0.6 m) length of size 20 fibrous wrapping twine. Similarly connect the other pad to a 1-ft (0.3 m) length of twine. Then securely connect the free ends of both lengths of twine to a small loop tied at the end of a similar 5-ft length of twine. The resulting assembly resembles the letter Y with a 5-ft long vertical stem and two diagonal branches (one 1 ft long and the other 2 ft long), with a folded swab securely attached at the end of each branch. After assembly, place each two-pad drag-swab sampling set into a sterile bag.

(2) Procedure for taking drag-swab.

(i) Floor litter: The Plan participants should collect two samples as follows: Drag four 3-by-3-inch sterile gauze pads premoistened with double strength skim milk over the floor litter surface for 15 min minimally. Place the gauze pads used to collect the samples in 18-oz whirl-pack bags, two pads per bag with each bag containing 5 ml of double strength skim milk. This will maintain the moistness of the sample during transport. Mark the bags with the type of sample and the house identification.

(ii) Nest-boxes. The Plan participant should collect one nest-box sample by using two 3-by-3-inch sterile gauze pads premoistened with double strength skim milk. Wipe the two gauze pads used to collect the sample over assorted locations of about 10 percent of the total nesting area. Place the gauze pads used to collect the sample in an 18-oz whirl-pack bag containing 5 ml of double strength skim milk. Mark the bag with the type of sample and the house identification.

(d) Chick-Papers

(i) Swabbing of Papers. Collect one chick box paper for each 10 boxes of chicks placed. Lay the chick papers on a clean surface. With latex-gloved hands, take a sterile 3-by-3 inch gauze pad saturated with double strength skimmed milk, and rub vigorously across the surface of the chick paper covering at least 75% of the area. Use sufficient pressure to rub any dry meconium off the papers. Pouring a small amount of milk (11 to 2 tablespoons) on each the paper will improve sample collection. Swab 5 chick papers per gauze pad. Place 2 gauze pads (a maximum of 10 combined swabbed papers) into an 18 oz Whirl-Pak bag, and add 1 to 2 tablespoons of skimmed milk. Gloves should be changes between each Whirl-Pak sample (each 10 papers), and at any time a glove is torn. Hands should be clean prior to swabbing and disinfectant should not be applied to the gloves. Transport samples on ice to a laboratory within 48 hours of collection. Samples can be frozen, for longer

storage, however immediate delivery to the laboratory is ideal.
(ii) Alternative to Swabbing Papers. An alternative to swabbing of chick papers at the farm or the delivery truck, is to send the chick papers directly to a laboratory. Select every 10th chick paper for culture. Place chick papers immediately into large plastic bags and close bags. Place bags in clean boxes and transport them in a timely manner to a laboratory. They do not need refrigeration.

Reason:
Chick papers is a very sensitive method for detecting the presence of Salmonella in chicks. These first chick droppings or meconium give a good indication of prior bacterial contamination. It was felt that since chick papers are used frequently to evaluate NPIP participating breeding flocks for salmonella that the collection protocol should be standardized.

Proponent:
General Conference Committee
National Poultry Improvement Plan

Proposal No. 20

Delegates:

B,C,E §147.11(a)(2) This proposal adds the collection of the bursa of Fabricius as part of the organ pool for bacteriological examination for salmonella.

(2) Selective enrichment culture (refer to illustration 2). Collect and culture organ samples separately from intestinal samples, with intestinal tissues collected last to prevent cross-contamination. Samples from the following organs or sites should be collected for culture in selective enrichment broth. A non-selective broth culture (illustration 1) of pooled organs and sites should also be included as described in paragraph (a)(3) of this section.

(i) Heart (apex, pericardial sac, and contents if present.);
(ii) Liver (portions exhibiting lesions or in grossly normal organs, the drained gallbladder and adjacent liver tissues.);
(iii) Ovary-Testes (entire inactive ovary or testes, but if ovary is active, include any atypical ova.);
(iv) Oviduct (if active, include any debris and dehydrated ova.);
(v) Kidneys and spleen;
(vi) Bursa of Fabricius (proximal wall); and,
(vii) Other visible pathological sites where purulent, necrotic, or proliferative lesions are seen.

(3) From each reactor, aseptically collect 10 to 15 g, or the nearest lesser amount available, from each organ or site listed in paragraph (a)(2) of this section and mince, grind, and blend them completely in 10 times their volume of beef extract broth or a comparable non-selective broth. Organs or sites listed in paragraph (a)(2) of this section may be pooled from the same individual bird. Suspensions should be transferred in 10-ml aliquots to 100ml of both tetrathionate brilliant green (TBG) (Hajna or Mueller-Kauffman) broth, and a separate non-selective broth and incubated 37°C for 24 hours. Refer to illustration 2 for recommended bacteriological recovery and identification procedures, *including delayed secondary enrichment* and combinations of plating media that significantly suppress the overgrowth of contaminants, such as brilliant green Novobiocin (BGN) and Xylose-Lysine-Tergitol 4 (XLT4).

(4) From each reactor, make a composite sample of the following parts of grossly normal or disease tissues from the digestive tract: Crop wall, duodenum (including portions of the pancreas), jejunum (including remnant of yolk-sac attachment), both ceca, cecal tonsils, and rectum-cloaca. Aseptically collect 10-15 g or the nearest lesser amount available from each specified digestive or intestinal tissue, and mince, grind, and blend them completely in 10 times their volume of TBG broth. The digestive/intestinal tissues may be pooled from the same individual bird. Do not pool tissues from different birds. Transfer 10 ml of the described digestive suspensions into 100 ml of TBG broth, and incubate at 41.5°C for 24 hours. Cultures may be incubated at 37°C if 41.5°C incubators are not available. The higher incubation temperatures for TBG broth reduce populations of competitive contaminants common in gut tissue. Refer to illustration 2 for recommended bacteriological recovery and identification procedures, *including delayed secondary enrichment* and combinations of plating media that significantly suppress the overgrowth of contaminants, such BGN and XLT4.

Reason:
The functions of the Bursa of Fabricius in the domestic fowl are not fully understood though-it is known to be involved in antibody production. Since the lumen of the Bursa communicates directly with the alimentary tract and its walls contain large aggregations of lymphocytes, it is possible that it may harbor common salmonella. Research has shown that salmonella were isolated more frequently from the Bursa of Fabricius than from any other site.

Proponent:
Dr. Ed Mallinson
Virginia-Maryland Regional College of Veterinary Medicine

Delegates:
B,C,E §147.11(a)(2) This proposal adds the collection of the bursa of Fabricius as part of the organ pool for bacteriological examination for salmonella.

(2) Selective enrichment culture (refer to illustration 2). Collect and culture organ samples separately from intestinal samples, with intestinal tissues collected last to prevent cross-contamination. Samples from the following organs or sites should be collected for culture in selective enrichment broth. A non-selective broth culture (illustration 1) of pooled organs and sites should also be included as described in paragraph (a)(3) of this section.

(i) Heart (apex, pericardial sac, and contents if present.);
(ii) Liver (portions exhibiting lesions or in grossly normal organs, the drained gallbladder and adjacent liver tissues.);
(iii) Ovary-Testes (entire inactive ovary or testes, but if ovary is active, include any atypical ova.);
(iv) Oviduct (if active, include any debris and dehydrated ova.);
(v) Kidneys and spleen;
(vi) Bursa of Fabricius (proximal wall); and,
(vii) Other visible pathological sites where purulent, necrotic, or proliferative lesions are seen.

(3) From each reactor, aseptically collect 10 to 15 g, or the nearest lesser amount available, from each organ or site listed in paragraph (a)(2) of this section and mince, grind, and blend them completely in 10 times their volume of beef extract broth or a comparable non-selective broth. Organs or sites listed in paragraph (a)(2) of this section may be pooled from the same individual bird. Suspensions should be transferred in 10-ml aliquots to 100ml of both tetrathionate brilliant green (TBG) (Hajna or Mueller-Kauffman) broth, and a separate non-selective broth and incubated 37°C for 24 hours. Refer to illustration 2 for recommended bacteriological recovery and identification procedures, *including delayed secondary enrichment* and combinations of plating media that significantly suppress the overgrowth of contaminants, such as brilliant green Novobiocin (BGN) and Xylose-Lysine-Tergitol 4 (XLT4).

(4) From each reactor, make a composite sample of the following parts of grossly normal or disease tissues from the digestive tract: Crop wall, duodenum (including portions of the pancreas), jejunum (including remnant of yolk-sac attachment), both ceca, cecal tonsils, and rectum-cloaca. Aseptically collect 10-15 g or the nearest lesser amount available from each specified digestive or intestinal tissue, and mince, grind, and blend them completely in 10 times their volume of TBG broth. The digestive/intestinal tissues may be pooled from the same individual bird. Do not pool tissues from different birds. Transfer 10 ml of the described digestive suspensions into 100 ml of TBG broth, and incubate at 41.5°C for 24 hours. Cultures may be incubated at 37°C if 41.5°C incubators are not available. The higher incubation temperatures for TBG broth reduce populations of competitive contaminants common in gut tissue. Refer to illustration 2 for recommended bacteriological recovery and identification procedures, *including delayed secondary enrichment* and combinations of plating media that significantly suppress the overgrowth of contaminants, such BGN and XLT4.

Reason:
Research has shown that Salmonella recovery has often been improved when the Bursa of Fabricius was also collected and cultured.
Proponent:
Dr. Ed Mallinson
Virginia-Maryland Regional College of Veterinary Medicine

Proposal No. 21

Delegates:

D §145.43(c) Amends this section by adding flock surveillance every 4-6 weeks.

(c) U.S. M. Gallisepticum Clean. (1) A flock maintained in accordance with the conditions and procedures described in § 147.26 of this chapter, and in which no reactors are found when a random sample of at least 10 percent of the birds in the flock, or 300 birds in flocks of more than 300 and each bird in flocks of 300 or less, is tested when more than 12 weeks of age, in accordance with the procedures described in § 145.14(b): Provided, That to retain this classification, a minimum of 30 samples from male flocks and 60 samples from female flocks shall be retested at 28-30 weeks of age and at 4-6 weeks thereafter.

(2) A flock qualified as U.S. M. Gallisepticum Clean may retain the classification through its first egg-laying cycle, provided it is maintained in isolation and no evidence of *M. gallisepticum* infection is revealed. A flock which is molted following completion of an egg-laying cycle and subsequently brought back into production, shall be retested within 2 weeks prior to production, as described in paragraph (c)(1) of this section. A State inspector shall visit with the owner or manager of each flock at least once during each laying cycle to discuss and ascertain whether the applicable conditions outlined in § 147.26 of this chapter are being met. If a flock proves to be infected with *M. gallisepticum*, it shall lose this classification.

(3) In order to sell hatching eggs or poults of this classification, all hatching eggs and poults handled by the participant must be of this classification.

Reason:
The provisions of the NPIP require that turkey breeding flocks be sampled at 4-6 week intervals to meet requirements for the U.S. MS and U.S. MM Clean classification under the Plan. Many States run the serum samples collected for MS and MM for MG. In an attempt to acquire uniformity, many States would like to see the same surveillance requirement for all Mycoplasmas.

Proponent:
Plan Staff, National Poultry Improvement Plan

Proposal No. 22

Delegates:
D §145.43(d) Amends the sample size for MM Clean qualification test.

(d) U.S. M. Meleagridis Clean. (1) A flock in which freedom from *M. meleagridis* has been demonstrated under the following criteria:
(i) A sample of ~~60~~ 100 birds from each flock has been tested for *M. meleagridis* when more than 12 weeks of age: Provided, That to retain this classification, a minimum of 30 samples from male flocks and 60 samples from female flocks shall be retested at 28-30 weeks of age and at 4-6 week intervals thereafter.

Reason:
U.S. MG Clean State, Turkeys are required to collect serum from 100 birds during the qualification test for MG. Usually the authorized laboratory runs a plate test for MG, MS, and MM on all the serum samples submitted. States have requested that the sample sizes for the qualification test and surveillance test be the same for all of the Plan Mycoplasmas.

Proponent:
Plan Staff, National Poultry Improvement Plan

Proposal No. 23

Delegates:
B,C,D,E §147.6 Amends this section as follows:

§ 147.6 Procedure for Determining the Status of Flocks Reacting to Tests for *Mycoplasma Gallisepticum*, *Mycoplasma Synoviae*, and ~~Mycoplasma~~ ~~Meleagridis~~.

The macroagglutination tests for Mycoplasma antibodies, as described in "Standard Methods for Testing Avian Sera for the Presence of Mycoplasma Gallisepticum Antibodies" published by the Agricultural Research Service, USDA, March 1966, and the microagglutination tests, as reported in the Proceedings, Sixteenth Annual Meeting of the American Association of Veterinary Laboratory Diagnosticians, 1973, shall be the official tests. Procedures for isolation and identification of Mycoplasma may be found in "Isolation and Identification of Avian Pathogens", published by the American Association of Avian Pathologists and §§ 147.15 and 147.16 of this part.

~~(a) When reactors are submitted to a laboratory as prescribed by the Official State Agency, the following criteria shall be used to determine if the flock is positive for M. gallisepticum, M. synoviae, or M. meleagridis:~~

~~(1) Active air sac lesions, sinusitis, synovitis, or other clinical signs of a respiratory disease;~~

~~(2) Recovery by culture of the Mycoplasma for which the flock was tested;~~

~~(3) Supplemental serological test.~~

~~(b) If all of these tests are negative, the flock shall be deemed to have had no reactors for the Mycoplasma for which the flock was tested. If the Mycoplasma for which the flock was tested is isolated bacteriologically or identified as infected by a polymerase chain reaction (PCR) based procedure approved by the Department, the flock shall be considered infected. If any of the other tests described in paragraphs (a) (1) or (3) of this section is positive, the flock shall be considered suspicious, and additional culturing procedures, and agglutination and hemagglutination inhibition (HI) tests shall be conducted according to the following sequence:~~

(a) The status of a flock for Mycoplasma shall be determined according to the following sequence and criteria:

(1) If the tube agglutination or the serum plate test is negative, the flock qualifies.

(2) If the tube agglutination or the serum plate test is positive, the hemagglutination inhibition (HI) test and/or the serum plate dilution (SPD) test shall be conducted.

(3) If the tube agglutination or serum plate tests are positive and HI and/ or the SPD tests are negative, the flock shall be retested in accordance with paragraph (~~b~~ a)(6) of this section.

(4) If HI titers of 1:40 or SPD titers of 1:5 are found, the flock shall be considered suspicious and shall be retested in accordance with paragraph (~~b~~ a)(6) of this section.

(5) If HI titers of 1:80, positive enzyme-labeled immunosorbent assay (ELISA) titers, or SPD titers of 1:10 or higher are found, ~~in conjunction with any of the criteria described in paragraph (a)(1) of this section,~~ the Official State Agency shall presume the flock to be infected. If the indicated titers are found, ~~but none of the criteria described in paragraph (a)(1) of this section are evident,~~ tracheal swabs from 30 randomly selected birds shall be taken promptly and cultured individually or a PCR-based procedure conducted on these specimens for Mycoplasma, and additional tests conducted in accordance with paragraph (~~b~~ a)(6) of this section before final determination of the flock status is made.

(6) Fourteen days after the previous bleeding date, all birds or a random sample comprised of 75 birds shall be tested by the serum plate or tube agglutination test. Tested birds shall be identified by numbered bands at the discretion of the Official State Agency.

(7) If the tube agglutination test or serum plate test is negative for the Mycoplasma for which the flock was tested, the flock qualifies.

(8) If the tube agglutination or serum plate test is positive, the HI and/

or SPD test shall be conducted on the reacting samples.

(9) On the retest, if the tube agglutination or serum plate tests are positive at the same or higher rate and the HI or SPD tests are negative, the flock shall be considered suspicious and shall be retested in accordance with paragraph (~~b~~ a)(6) of this section.

(10) On the retest if HI titers of 1:80 and/or SPD titers of 1:10 or higher are found, the flock shall be considered infected: Provided, That, at the discretion of the Official State Agency, additional tests may be conducted in accordance with paragraph (~~b~~ a)(6) of this section before final determination of the flock status is made.

(11) If HI titers of 1:80 and/or SPD titers of 1:10 or higher are found on the second retest, the flock shall be considered infected for the Mycoplasma for which it was tested.

(12) If the tube agglutination or serum plate tests are found on the second retest to be positive at the same or higher rate and the HI and/or SPD tests are negative, the flock should be considered infected: Provided, That if the status of the flock is considered to be equivocal, the Official State Agency may examine reactors by the in vivo bio-assay, PCR-based procedures, and/or culture procedures before final determination of the flock status is made.

(13) If the in vivo bio-assay, PCR-based procedures, and culture procedures are both negative, the Official State Agency may qualify the flock for the classification for which it was tested.

(14) If the in vivo bio-assay, PCR-based procedures, or culture procedures are positive, the flock shall be considered infected: Provided, That if only the bio-assay is positive, additional in vivo bio-assay, PCR-based procedures, or cultural examinations may be conducted by the Official State Agency before final determination of the flock status is made.

(15) If the in vivo bio-assay, PCR-based procedures, or cultures are positive on retest, the flock shall be considered infected for the Mycoplasma for which it was tested.

Reason:
Authorized laboratories and Official State Agencies of the NPIP conduct the serum plate test or the ELISA as the screening test for the Mycoplasmas. The HI is conducted on the plate reactors or ELISA suspects. If HI titers of 1:80, positive ELISA titers or SPD titers of 1:10 or higher are found, confirmatory culture or PCR-based procedure tests are conducted to determine the status of the flock.

Proponent:
Dr. Louis van der Heide
University of Connecticut
Storrs, Connecticut

)elegates:
B,C §147.11 amends this section by adding the option of using a rapid Salmonella enteritidis test in lieu of the recommended culture protocol.

§ 147.11 Laboratory Procedure Recommended for the Bacteriological Examination of Salmonella.

(a) For egg- and meat-type chickens, waterfowl, exhibition poultry, and game birds. All reactors to the Pullorum-Typhoid tests, up to at least four birds, should be cultured in accordance with both direct (paragraph (a)(1)) and selective enrichment (paragraph (a)(2)) procedures described in this section. Careful aseptic technique should be used when collecting all tissue samples. A 24 hour micro-magnetic bead Salmonella screen with a monoclonal antibody latex bead confirmation test for *Salmonella enteritidis* may be used in lieu of the direct culture and/or selective enrichment culture methods described in § 147.11 of this chapter for egg-type breeding flocks participating in the U.S. *S. enteritidis* Monitored program and meat-type breeding flocks participating in the U.S. *S. enteritidis* Clean program.
(a) Except when visibly pathological tissues are present, direct culture, § 147.11(a)(1) of this subpart, may be omitted; and
(b) Enrichment culture of organ (non-intestinal) tissues using a non-selective broth, § 147.11(a)(2) of this subpart, may be omitted.

Reason:
The General Conference Committee approved on an interim basis a 24 hour micro-magnetic bead Salmonella screen with a monoclonal antibody confirmation for Salmonella enteritidis. Such interim approval shall remain in effect until confirmed or rejected by the next Plan Conference, or until rescinded by the GCC. Additional field trials comparing the rapid test with the conventional salmonella culture methods of the NPIP have been conducted to provide scientific information for the technical committee and the Official delegates when they consider final approval for the rapid test at the Plan Conference.

Proponent:
General Conference Committee

Delegates:
B,C,D §§145.23(d), 145.33(d), 145.43(f) adds the use of only FDA approved anti-salmonella additives for feed in.

(d) U.S. S. Enteritidis Monitored. This program is intended to be the basis from which the breeding-hatching industry may conduct a program for the prevention and control of Salmonellosis. It is intended to reduce the incidence of Salmonella organisms in hatching eggs and chicks through an effective and practical sanitation program at the breeder farm and in the hatchery. This will afford other segments of the poultry industry an opportunity to reduce the incidence of Salmonella in their products.
(1) A flock and the hatching eggs and chicks produced from it which have met the following requirements as determined by the Official State Agency:
(i) The flock originated from a U.S. Sanitation Monitored flock, or meconium from the chick boxes and a sample of chicks that died within 7 days after hatching are examined bacteriologically for salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped.
(ii) All feed fed to the flock shall meet the following requirements:
(A) Pelletized feed shall contain either no animal protein or only animal protein products produced under the Animal Protein Products Industry (APPI) Salmonella Education/Reduction Program[2]. The protein products must have a minimum moisture content of 14.5 percent and must have been heated throughout to a minimum temperature of 190 °F, or above, or to a minimum temperature of 165 °F. for at least 20 minutes, or to a minimum temperature of 184 °F. under 70 lbs. pressure during the manufacturing process.
(B) Mash feed shall contain either no animal protein or only animal protein products supplement manufactured in pellet form and crumbled.
(C) Only FDA approved products that prevent Salmonella contamination of feed may be used.

Reason:
This change would provide additional tools that may be used to insure salmonella free feed for a breeding flock that participates in U.S. *S. enteritidis* Monitored program for egg-type chickens, U.S. *S. enteritidis* Clean for primary meat-type chickens, and U.S. Sanitation Monitored program for both meat-type chickens and turkeys. salmonella in a poultry breeding flock.

Proponent:
Dr. Douglas Waltman III
Oakwood, Georgia

[2]Documents concerning the APPI/Salmonella Education Reduction Program may be obtained from Mr. A. R. Rhorer; USDA,APHIS,VS, Suite A-102, 1500 Klondike Road, Conyers, Georgia 30207.

Delegate
D *§145.43(f) U.S. Sanitation Mnnitored, Turkeys is amended by deleting paragraph(f)(7).*

(f) U.S. Sanitation Monitored, Turkeys. A flock or hatchery whose owner is controlling or reducing the level of salmonella through compliance with sanitation and management practices as described in Subpart C of Part 147 of this chapter, and where the following monitoring, testing, and management practices are conducted:

(1) Hatchery debris (dead germ hatching eggs, fluff, and meconium collected by sexors), a sample of the poults that died within 10 days after hatching, or both, from each candidate breeding flock produced by a primary breeder, are examined bacteriologically at an authorized laboratory for Salmonella.

(2) The poults for the candidate breeding flock are placed in a building that has been cleaned, disinfected, and examined bacteriologically for the presence of Salmonella by an Authorized Agent, as described in sec. 147.12 of this chapter.

(3) Feed for turkeys in the candidate breeding flock shall meet the following requirements:

(i) All feed manufactured in pellet form must contain a minimum moisture content of 14.5 percent and must have been heated throughout to a minimum temperature of 190 °F. or above, or to a minimum temperature of 165 °F. for at least 20 minutes, or to a minimum temperature of 184 °F. under 70 lbs. pressure during the manufacturing process.

(ii) Initial feed (for newborn poults to 2 weeks of age) shall be manufactured in pellet form, either with no animal protein or with animal protein products produced under the Animal Protein Products Industry Salmonella Education/ Reduction Program.

(iii) Succeeding feed (for turkeys 2 weeks or older) shall be as described in (f)(3)(ii) of this section, mash that contains no animal protein products, or mash that contains an animal protein products supplement that has been manufactured in pellet form and crumbled.

(4) Environmental samples shall be taken by an Authorized Agent, as described in sec. 147.12 of this chapter, from each flock at 12-20 weeks of age and examined bacteriologically at an authorized laboratory for Salmonella.

(5) Owners of flocks found infected with a paratyphoid Salmonella may vaccinate these flocks with an autogenous bacterin with a potentiating agent. /6/

NOTE /6/ Preparation and use of this type of vaccine may be regulated by state statutes.

(6) Environmental samples shall be taken by an Authorized Agent, as described in sec. 147.12 of this chapter, from each flock at 35-50 weeks of age and from each molted flock at midlay, and examined bacteriologically at an authorized laboratory for Salmonella.

~~(7) Environmental samples shall be taken, by an Authorized Agent using the procedures described in sec. 147.12 of this chapter, from the laying house after the flock is removed, and examined bacteriologically at an authorized laboratory for Salmonella.~~

(8) Hatchery debris (dead germ hatching eggs, fluff, and meconium collected by sexors), a sample of the poults that died within 10 days after hatching, or both shall be cultured from poults produced from hatching eggs from each fl ock, as a means of evaluating the effectiveness of the control procedures.

Reason:
The required activity of this paragraph adds cost to the program but little else. No real information or answers are to come from this, after the fact, testing. Yes it might confirm the presence of a

potential Pathogen(s) in the litter but would not necessarily be an indication the same pathogen was or was not present in the flock. If the flock was already marketed it would be too late to take any effective action anyway. Neither would such a test be a relevant indicator for the next flock in the facility since it would be done prior to cleanup and disinfection.

Proponent:
Mr. Bill Mattos
California Poultry Health Board

Proposal No. 27

Delegates
B,C,D,E,F § 147.42 Amends this section

sec. 147.42 General.
Changes in this subchapter shall be made in accordance with the procedure described in this subpart: Provided, That the Department reserves the right to make changes in this subchapter without observance of such procedure when such action is deemed necessary in the public interest:Provided further, that the department give the General Conference Committee 60 days'written notice to the changes in this subchapter.

Reason:
Changes to provisions of the NPIP (a voluntary program) should not be made with due discussion with and input from the GCC, the State Organizations and any and all interested Plan participants.

Proponent:
Mr. Bill Mattos
California Poultry Health Board

Delegates

E §145.53(d) Creates a Salmonella enteritidis monitored program for subpart E participants.

(d) U.S. S. Enteritidis Monitored. This program is intended to be the basis from which the breeding-hatching industry may conduct a program for the prevention and control of Salmonellosis. It is intended to reduce the incidence of Salmonella organisms in hatching eggs and chicks through an effective and practical sanitation program at the breeder farm and in the hatchery. This will afford other segments of the poultry industry an opportunity to reduce the incidence of Salmonella in their products.
(1) A flock and the hatching eggs and chicks produced from it which have met the following requirements as determined by the Official State Agency:
(i) The flock originated from a U.S. Sanitation Monitored flock, or meconium from the chick boxes and a sample of chicks that died within 7 days after hatching are examined bacteriologically for salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped.
(ii) All feed fed to the flock shall meet the following requirements
(A) Pelletized feed shall contain either no animal protein or only animal protein products produced under the Animal Protein Products Industry (APPI) Salmonella Education/Reduction Program[3]. The protein products must have a minimum moisture content of 14.5 percent and must have been heated throughout to a minimum temperature of 190 °F, or above, or to a minimum temperature of 165 °F. for at least 20 minutes, or to a minimum temperature of 184 °F. under 70 lbs. pressure during the manufacturing process.
(B) Mash feed shall contain either no animal protein or only animal protein products supplement manufactured in pellet form and crumbled.
(iii) Feed shall be stored and transported in such a manner as to prevent possible contamination;
(iv) The flock is maintained in compliance with sec.sec. 147.21, 147.24(a), and 147.26 of this chapter;
(v) Environmental samples shall be collected from the flock by an Authorized Agent, as described in sec. 147.12 of this chapter, when the flock is 2 to 4 weeks of age. The Authorized Agent shall also collect samples every 30 days after the first sample has been collected. The samples shall be examined bacteriologically for group D salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped.
(vi) A federally licensed Salmonella enteritidis bacterin may be used in multiplier breeding flocks that are negative for Salmonella enteritidis upon bacteriological examination as described in paragraph (d)(1)(v) of this section: Provided, that a sample of 350 birds, which will be banded for identification, shall remain unvaccinated until the flock reaches at least 4 months of age. Following negative serological and bacteriological examinations as described in paragraph (d)(1)(vii) of this section, the banded, non-vaccinated birds shall be vaccinated.
(vii) Blood samples from 300 non-vaccinated birds as described in paragraph (d)(1)(vi) of this section shall be officially tested with pullorum-typhoid antigen when the flock is a minimum of more than 4 months of age. All birds with positive or inconclusive reactions, up to a maximum of 25 birds, shall be submitted to an authorized laboratory and examined for the presence of group D salmonella, as described in sec. 147.11 of this chapter. Cultures from positive samples shall be serotyped.

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[3]Documents concerning the APPI/Salmonella Education Reduction Program may be obtained from Mr. A. R. Rhorer; Sheep, Goat, Equine, and Poultry Diseases Staff; VS, APHIS, USDA; Room 205 Presidential Building; 6525 Belcrest Road; Hyattsville, Maryland 20782.

(viii) Hatching eggs are collected as quickly as possible and are handled as described in sec. 147.22 of this chapter and are sanitized or fumigated (see sec. 147.25 of this chapter).
(ix) Hatching eggs produced by the flock are incubated in a hatchery that is in compliance with the recommendations in sec.sec. 147.23 and 147.24(b) of this chapter, and sanitized either by a procedure approved by the Official State Agency or fumigated (see sec. 147.25 of this chapter).
(2) A flock shall not be eligible for this classification if Salmonella enteritidis ser enteritidis (SE) is isolated from a specimen taken from a bird in the flock. Isolation of SE from an environmental or other specimen as described in section (d)(1)(v) of this paragraph will require bacteriological examination for SE in an authorized laboratory, as described in sec. 147.11(a) of this chapter, of a random sample of 60 live birds from a flock of 5,000 birds or more, or 30 live birds from a flock with fewer than 5,000 birds. If only one specimen is found positive for SE, the participant may request bacteriological examination of another 60-bird sample from the flock. If no SE is recovered from any of the specimens in the second sample, the flock will be eligible for the classification.
(3) A flock shall be eligible for this classification if Salmonella enteritidis (S. enteritidis ser Enteritidis) is isolated from an environmental sample collected from the flock in accordance with paragraph (d)(v) of this section: Provided, That testing is conducted in accordance with paragraph (d)(1)(vi) of this section each 30 days and no positive samples are found.
(4) In order for a hatchery to sell products of this classification, all products handled shall meet the requirements of the classification.
(5) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures.

Reason:
This would allow those subpart E participants to hatch waterfowl, exhibition poultry and game birds in the same hatchery with egg and meat-type chicken flocks participating in a *Salmonella enteritidis* program of the NPIP.

Proponent:
Mr. Paul Brennan
Indiana State Poultry Association

Proposal No. 29

Delegates:
E §145.53(e) Adds a Mycoplasma synoviae Clean program for subpart E participants:

(e) U.S. M. Synoviae Clean. (1) A flock maintained in compliance with the provisions of sec. 147.26 and in which freedom from M. synoviae has been demonstrated under the criteria specified in paragraph (e)(1) (i) or (ii) of this section.
(i) It is a flock in which a minimum of 300 birds has been tested for M. synoviae as provided in sec. 145.14(b) when more than 4 months of age: Provided, That to retain this classification, a sample of at least 150 birds shall be tested at intervals of not more than 90 days: And provided further, That a sample comprised of less than 150 birds may be tested at any one time, with the approval of the Official State Agency and the concurrence of the Service, provided that a minimum of 150 birds is tested within each 90-day period; or
(ii) It is a multiplier breeding flock which originated as U.S. M. synoviae Clean chicks from primary breeding flocks and from which a sample comprised of a minimum of 75 birds has been tested for M. synoviae as provided in sec. 145.14(b) when more than 4 months of age: Provided, That to retain this classification, the flock shall be subjected to one of the following procedures:
(A) At intervals of not more than 90 days, a sample of 50 birds shall be tested: Provided, That a sample of less than 50 birds may be tested at any one time, provided that a minimum of 30 birds per flock with a minimum of 15 birds per pen, whichever is greater, is tested each time and a total of at least 50 birds is tested within each 90-day period; or
(B) At intervals of not more than 30 days, egg yolk testing shall be conducted in accordance with sec. 147.8.
(2) A participant handling U.S. M. Synoviae Clean products shall keep these products separate from other products in a manner satisfactory to the Official State Agency: Provided, That U.S. M. Synoviae Clean chicks from primary breeding flocks shall be produced in incubators and hatchers in which only eggs from flocks qualified under paragraph (e)(1)(i) or (ii) of this section are set.
(3) U.S. M. synoviae Clean chicks shall be boxed in clean boxes and delivered in trucks that have been cleaned and disinfected as described in sec. 147.24(a).

Reason:
This would allow those subpart E participants to hatch waterfowl, exhibition poultry and game birds in the same hatchery with egg and meat-type chicken flocks participating in a *Mycoplasma synoviae* program of the NPIP.

Proponent:
Mr. Paul Brennan
Indiana State Poultry Association